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PATENT

**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

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	)	
King et al.	)	Art Unit: 1643
	)	
Application No. 07/110,791	)	Examiner: Stephen L. Rawlings
	)	
Filing Date: October 21, 1987	)	Confirmation No. 7373
	)	
For: HUMAN GENE RELATED TO BUT	)	
DISTINCT FROM EGF RECEPTOR	)	
GENE	)	

**DECLARATION OF C. RICHTER KING, MATTHIAS H. KRAUS AND  
STUART A. AARONSON UNDER 37 C.F.R. § 1.131**

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Sir:

We, C. RICHTER KING, MATTHIAS H. KRAUS and STUART A. AARONSON

declare as follows:

1. We are the applicants of the above-identified patent application and co-inventors of the subject matter described and claimed therein.

2. Prior to October, 1985 and January, 1986, we reduced the claimed invention to practice as evidenced by our attached publication, King et al., *Science* 229:974-976 (September, 1985) (submitted as an Exhibit in the previously filed Declaration by us). As indicated on page 976, our manuscript was submitted to the editors of the journal, *Science* on April 22, 1985 and accepted on June 3, 1985. The inventive activities embodied in the King et al. publication all took place in the United States.

3. We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements are made with the knowledge that willful false statements and the like are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that any such false statement may jeopardize the validity of the application or any patent issued thereon.

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ORIGINAL  
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501  
1 Title: A HUMAN GENE RELATED TO BUT DISTINCT  
2 FROM EGF RECEPTOR GENE

3 *Insert E1* Background of the Invention

4 Technical Field

5 The present invention is related to the cloning,  
6 isolation and partial characterization of a hitherto  
7 unidentified human gene. More particularly, the present  
8 invention is related to the preparation and  
9 identification of a v-erbB related human gene that is a  
10 new member of the tyrosine kinase encoding family of  
11 genes and is amplified in a human mammary carcinoma.

12 State of the Art

13 A number of genes have been identified as retroviral  
14 oncogenes that are responsible for inducing tumors in  
15 vivo and transforming cells in vitro (Land et al.,  
16 Science 222:771-778, 1983). Some of them apparently  
17 encode transforming proteins that share a kinase domain

1 homologous to that of pp60<sup>src</sup>, a tyrosine-specific protein  
2 kinase. The cellular cognate, encoded by the c-src gene,  
3 also exhibits tyrosine-specific kinase activity. Of  
4 particular interest is the fact that tyrosine-specific  
5 kinases are also encoded by other genes for  
6 several receptors for polypeptide growth factors,  
7 including the receptors for epidermal growth factor (EGF)  
8 (Cohen et al., J. Biol. Chem. 255:4834-4842, 1980),  
9 platelet-derived growth factor (PDGF) (Nishimura et al.,  
10 Proc. Natl. Acad. Sci. USA 79:4303-4307, 1982), insulin  
11 (Kasuga et al., Nature 298:667-669, 1982), and  
12 insulin-like growth factor I (Rubin et al., Nature  
13 305:438-440, 1983). This implies a possible link between  
14 the action of the growth factor-receptor<sup>7</sup> complex and the  
15 oncogene products with tyrosine-specific kinase activity.

16 Recent analysis of the v-erbB gene and the EGF  
17 receptor gene indicates that the v-erbB gene is a part of  
18 the EGF receptor gene and codes for the internal domain  
19 and transmembrane portion of the receptor (Yamamoto et  
20 al., Cell 35:71-78, 1983; Downward et al., Nature  
21 307:521-527, 1984; Ullrich et al., Nature 309:418-425,  
22 1984). These findings, together with the extensive  
23 identity of the amino acid sequences of the v-sis protein  
24 and platelet-derived growth factor (Waterfield et al.,

1 Nature 304:35-39, 1983; Doolittle et al., Science  
2 221:275-277, 1983), suggest that some viral oncogene  
3 products mimic the action of the polypeptide growth  
4 factor-receptor complex in activating a cellular pathway  
5 involved in cell proliferation and tumor formation.

6 Genetic alterations affecting proto-oncogenes of the  
7 tyrosine kinase family may play a role in spontaneous  
8 tumor development. A specific translocation affecting  
9 the c-abl locus, for example, is associated with chronic  
10 myelogenous leukemia (de Klein et al., Nature 300:765,  
11 1982; Collins et al., Proc. Natl. Acad. Sci. USA 80:4813,  
12 1983). Several recent studies have also documented  
13 amplification or rearrangement of the gene for the EGF  
14 receptor in certain human tumors (Libermann et al.,  
15 Nature 313:144, 1985), or tumor cell lines (Ullrich et  
16 al., Nature 309:418, 1984; Lin et al., Science 224:843,  
17 1984). However, a gene that is a new member of the  
18 tyrosine kinase family and is amplified in a human  
19 mammary carcinoma and is closely related to, but distinct  
20 from the EGF receptor gene, has not heretofore been known.

SUMMARY OF THE INVENTION

It is, therefore, an object of the present invention to provide a novel, cloned, human gene having the nucleotide sequence as shown in Fig. 1 and described more fully herein infra.

It is a further object of the present invention to provide products, e.g. various RNAs and/or polypeptides encoded by the cloned gene.

It is a still further object of the present invention to provide antibodies, either polyclonal or monoclonal, directed against the protein product encoded by said gene and a diagnostic kit containing said antibodies for the detection of carcinomas.

It is another object of the present invention to provide complementary DNA (cDNA) clones homologous to the messenger RNA (mRNA) encoded by the cloned gene, said cDNA clones being capable of expressing large amounts of corresponding protein in a heterologous vector system, such as bacteria, yeast, and the like.

*eukaryotes*

It is yet another object of the present invention to produce a transformed cell or organism capable of

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1     expressing said gene by incorporating said gene or a part  
2     thereof into the genome of said cell, vector or organism.

3             It is a still further object of the present  
4     invention to provide nucleic acid probes and/or antibody  
5     reagent kits capable of detecting said gene or a product  
6     thereof.

7             Other objects and advantages of the present  
8     invention will become apparent as the detailed  
9     description of the invention proceeds.

10                     BRIEF DESCRIPTION OF DRAWINGS

11             These and other objects, features and many of the  
12     attendant advantages of the invention will be better  
13     understood upon a reading of the following detailed  
14     description when considered in connection with the  
15     accompanying drawings wherein:

16             ~~Fig. 1 shows a characteristic fragment produced by~~  
17     ~~EcoRI restriction of the cloned gene of the present~~  
18     ~~invention; detection of v-erbB and pMAC 117 specific~~  
19     ~~gene fragments in normal human placenta, A431 cells, or~~  
20     ~~human mammary carcinoma MAC117. DNA (15 pg) was cleaved~~  
21     ~~with Eco RI, separated by electrophoresis in agarose~~

*sub*  
*Q*



1 gels, and transferred to nitrocellulose paper (Southern  
2 J. Mol. Biol. 98:503, 1975). Hybridization to the  
3 <sup>32</sup>P-labeled probe (Rigby et al., J. Mol. Biol. 113:237,  
4 1977) was conducted in a solution of 40 percent  
5 formamide, 0.75M NaCl, 0.075M sodium citrate, at 42°C  
6 (Wahl et al., Proc. Natl. Acad. Sci. USA 76:3683, 1979).  
7 The v-erbB probe (A) was a mixture of the 0.5-kbp Bam  
8 HI-Bam HI fragment and 0.5-kbp Bam HI-Eco RI fragment of  
9 avian erythroblastosis proviral DNA. The pMAC117 probe  
10 (B) was a 1-kbp Bgl I-Bam HI fragment. After  
11 hybridization, the blots were washed first in 0.3M NaCl  
12 plus 0.03M sodium citrate at room temperature, and then  
13 in 0.015M NaCl, 0.0015M sodium citrate, and 0.1 percent  
14 sodium dodecyl sulfate at 42°C (A) or at 52°C (B).  
15 Hybridization was detected by autoradiography.

16 Fig. 2 shows the gel electrophoretic properties of  
17 specific gene fragments; Restriction-site map of MAC117  
18 and plasmid pMAC117. A, Acc I; B, Bam HI; Bg, Bgl I; N,  
19 Nco I; R, Eco RI; X, Xba I; Xh, Xho I. The sites were  
20 located by electrophoretic analysis of the products of  
21 single and double digestion. Regions homologous to  
22 v-erbB or human repetitive sequences (region flanked by  
23 arrows) were located by Southern blot hybridization

1 (Southern, J. Mol. Biol. 98:503, 1975) with the v-erb B  
2 probe or total human DNA made radioactive by nick  
3 translation (Rigby et al., J. Mol. Biol. 113:237, 1977).  
4 Hybridization conditions were as described in Fig. 1A.  
5 The nucleotide sequence of pMAC117 between the Acc I site  
6 and the Nco I sites and regions of encoded amino acid  
7 sequence homologous to the EGF receptor are shown. The  
8 AG or GT dinucleotides flanking the putative coding  
9 regions are underlined. To determine the sequence, Nco  
10 I, Hinf I, and Sau 96 I fragments were labeled at the 3'  
11 termini by means of the large fragment of E. coli DNA  
12 polymerase, separated into single strands by gel  
13 electrophoresis, and chemically degraded (Maxam et al.,  
14 Proc. Natl. Acad. Sci. USA 74:560, 1977).

15 Fig. 3 shows a comparison of the putative encoded  
16 amino acid sequence of various polypeptide products, and  
17 comparison of the putative encoded amino acid sequence in  
18 pMAC117 with known tyrosine kinase sequences. Black  
19 regions represent homologous amino acids. Differing  
20 amino acid residues are shown in one-letter code (A,  
21 alanine; C. cysteine, D. aspartic acid; E. glutamic acid;  
22 F. phenylalanine; G. glycine; H. histidine; I.  
23 isoleucine; K. lysine; L. leucine; M. methionine;

1 N. asparagine; P. proline; Q. glutamine; R. arginine; S.  
2 serine; T. threonine; V. valine; W. tryptophan; Y.  
3 tyrosine). Amino acid positions conserved in all  
4 sequences are denoted by \*. The tyrosine homologous to  
5 that autophosphorylated by the v-src protein (Smart et  
6 al., Proc. Natl. Acad. Sci. USA 78:6013, 1981) is shown  
7 by an arrow. The v-abl sequence contains a tyrosine  
8 residue in this region displaced by two positions. The  
9 amino acid sequences of human EGF receptor, v-src, v-abl,  
10 v-fms, and human insulin receptor were aligned by the  
11 computer program described by Ullrich et al., Nature  
12 313:756, 1985 which is incorporated herein by reference.  
13 The homology observed with the predicted amino acid  
14 sequences of v-yes and v-fes was 51 percent and 48  
15 percent, respectively.

16 Fig. 4 shows the distinction between  $\lambda$ MAC117 and  
17 human EGF receptor genes by the  
18 detection of distinct messenger RNA species derived from  
19 the  $\lambda$ MAC117 gene and the human EGF receptor gene.  
20 Polyadenylated messenger RNA of A431 cells was separated  
21 by denaturing gel electrophoresis in formaldehyde  
22 (Lehrach et al., Biochemistry 16:4743, 1977), transferred  
23 to nitrocellulose (Southern, J. Mol. Biol. 98:503, 1975),  
24 and hybridized under stringent conditions (50 percent  
25 formamide, 0.75 M NaCl, 0.075M sodium citrate, at 42°C)

8  
1 with  $^{32}\text{P}$ -labeled probe from pMAC117 (Bgl I-Bam HI  
2 fragment) or human EGF receptor complementary DNA <sup>pE7s</sup>~~(pE7)~~  
3 2-kb Cla I inserted fragment). Filters were washed under  
4 conditions of high stringency (0.015M NaCl plus 0.0015M  
5 sodium citrate at 55°C). Hybridization was detected by  
6 autoradiography with exposure times of 4 hours for the  
7 pMAC117 probe and 1 hour for the human EGF receptor  
8 probe.

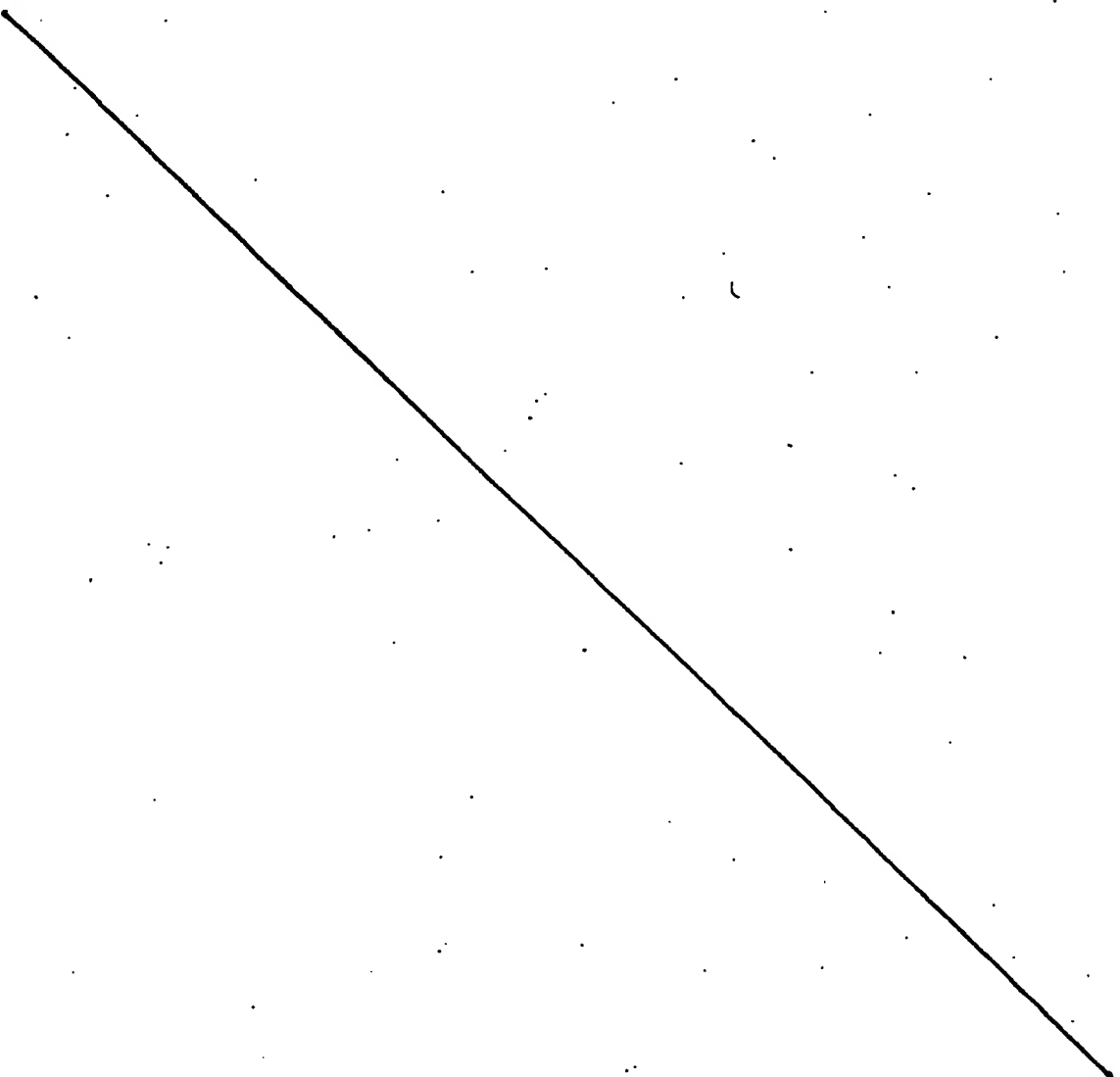


Fig. 5A shows the restriction map of complementary DNA of MAC117 encompassing the entire coding region of the gene. Clone pMAC137 was isolated from an oligo dT primed normal human fibroblast cDNA library (Okuyama et al., Mol. Cell. Biol. 3, 280, 1983) using a 0.8-kbp Acc I fragment from <sup>the 3' terminus of</sup> pMAC117 as probe. Clones  $\lambda$ MAC30,  $\lambda$ MAC10', and  $\lambda$ MAC14-1 were subsequently isolated from a randomly primed MCF-7 cDNA library (Walter et al., Proc. Natl. Acad. Sci. USA, 82, 7889, 1985) using cDNA fragments as probes. Restriction sites: B - Bam HI, BII - Bst EII, E - Eco RI, N - Nco I, P - Pst I, Sm - Sma I, Sp - Sph I, and St - Stu I.

Fig. 6 shows the overexpression of MAC117 in RNA in human mammary tumor cell lines. (A) Northern blot analysis. Total cellular RNA (10  $\mu$ g) of mammary tumor cell lines, normal fibroblasts M413 and HBL100 was hybridized with a cDNA probe derived from the 5' end of the coding region (Fig. 1B, probe a). M413 and HBL100 cells contain specific mRNA detectable after longer autoradiographic exposures. Similar results were obtained when probe b or c (Fig. 5B) was employed for hybridization. (B) Quantitation of mRNA levels. Serial 2-fold dilutions of total RNA were applied to nitrocellulose. Replicate filters were hybridized with either a cDNA probe (Fig. 1B, probe b) or human  $\beta$ -actin which served as control for RNA amounts present on the nitrocellulose filter. Relative amounts detected with each probe are indicated in comparison to the hybridization signals observed in normal human fibroblast M413.

Fig. 7 shows the 185-kDa protein specific for MAC117 and its overexpression in human mammary tumor cell lines. 40  $\mu$ g total cellular protein was separated by electrophoresis and transferred to nitrocellulose filters. The protein was detected with an antipeptide antibody coupled to  $^{125}$ I protein A. The

specificity of antibody detection was determined by pre-incubation of the antibody with excess amounts of peptide prior to immunodetection.

(+) preincubation with peptide, (-) no peptide. In panel B, nonspecific bands at 100 kd are observed in longer exposures of peptide blocked immunoblots (panel A).

Fig. 8 shows the gene amplification of MAC117 in 4 mammary tumor cell lines and the absence of MAC117 gene amplification in 4 other mammary tumor cell lines overexpressing MAC117 mRNA. (A) Southern blot analysis. For each line 10 µg genomic DNA were restricted with Xba I and hybridized with a probe comprising the entire coding region of MAC117 (Fig. 5B, probe c). Hind III restriction fragments of lambda DNA served as mol. wt. standards. (B) DNA dot-blot analysis. Genomic DNA (10 µg) digested with Eco RI was applied in serial 2-fold dilutions to nitrocellulose filters. Filters were hybridized either with a probe specific for MAC117 (Fig. 5B, probe b) or mos, which served as a control for DNA amounts applied to replicate nitrocellulose filters. Gene copy numbers of MAC117 relative to M413 indicate the minimal extent of gene amplification detected in DNA from mammary tumor cell lines.


Fig. 9 depicts the construction of expression vectors for the human MAC117 cDNA. A Nco I-Mst II fragment encompassing the entire open reading frame was cloned under the transcriptional control of either the SV40 early promoter or MuLV LTR. Symbols: , erbA-erbB intergenic region of pAEV11 containing the 3' splice acceptor site; N = Nco I, Sp = Sph I, M = Mst II, St = Stu I, H = Hind III, Sm = Sma I, P = Pst I, B = BamH I, X = Xho I. Sites indicated in parenthesis were not reconstituted after the cloning procedures.

Table 1 compares transformation characteristics of NIH/3T3 cells transfected with vectors generating different expression levels of the MAC117 coding sequence.

Table 1.

DNA transfectant <sup>a</sup>	Specific transforming activity <sup>b</sup> (ffu/pM)	Colony-forming efficiency in agar (%) <sup>c</sup>	Cell number required for 50% tumor incidence <sup>d</sup>
LTR-1/MAC117	$4.1 \times 10^4$	45	$10^3$
SV40/MAC117	$<10^0$	$<0.01$	$>10^6$
LTR/erbB	$5.0 \times 10^2$	20	$5 \times 10^4$
LTR/ <u>ras</u>	$3.6 \times 10^4$	35	$10^3$
pSV2/gpt	$<10^0$	$<0.01$	$>10^6$

<sup>a</sup>All transfectants were isolated from plates which received 1  $\mu$ g cloned DNA and were selected by their ability to grow in the presence of killer HAT medium (Mulligan et al., Proc. Natl. Acad. Sci. USA 78, 2072, 1981).

<sup>b</sup>Focus-forming units were adjusted to ffu/pM of cloned DNA added based on the relative molecular weights of the respective plasmids.

<sup>c</sup>Cells were plated at 10-fold serial dilutions in 0.33% soft agar medium containing 10% calf serum. Visible colonies comprising  $>100$  cells were scored at 14 days.

<sup>d</sup>NFR nude mice were inoculated subcutaneously with each cell line. Ten mice were tested at cell concentrations ranging from  $10^6$  <sup>to  $10^3$</sup>   ~~$10^6$  to  $10^3$~~  cells/mouse. Tumor formation was monitored at least twice weekly for up to 30 days.

j Fig. 10 shows the comparison of the levels of MAC117 proteins in LTR-1/erbB-2 transformed NIH/3T3 cells and human mammary tumor lines by immunoblot analysis. Varying amounts of total cellular protein ~~was~~<sup>MAC117</sup> separated by electrophoresis and transferred to nitrocellulose filters. The MAC117 protein was detected with rabbit anti-peptide serum coupled to <sup>125</sup>I protein A as previously described.

9

#### DETAILED DESCRIPTION OF INVENTION

10           The above and other objects and advantages of the  
11   present invention are achieved by a cloned human gene  
12   having the nucleotide sequence as shown in Fig. 1.  
13   Although any methods and materials similar or equivalent  
14   to those described herein can be used in the practice or  
15   testing of the present invention, the preferred methods  
16   and materials are now described. All publications  
17   mentioned under the "Brief Description of Drawings" and  
18   hereunder are incorporated herein by reference. Unless  
19   defined otherwise, all technical or scientific terms used  
20   herein have the same meaning as commonly understood by  
21   one of ordinary skill in the art to which this invention  
22   belongs.



1     Cells and Tissues:

2             Preparation of High Molecular Weight DNA

3             1.   From A431 cells:

4             A431 carcinoma cells were established in culture and  
5             maintained in Dulbecco's modified Eagle's medium with 10%  
6             fetal calf serum.

7             Cells were grown to 90% confluence in four 175 cm<sup>2</sup>  
8             tissue culture flasks, washed twice with phosphate  
9             buffered saline (Gibco Biochemicals), then lysed in 10 mM  
10            Tris (pH 7.5), 150 mM NaCl, 50 mM ethylenediamine-  
11            tetraacetate (EDTA) and 0.5% sodium dodecyl sulfate  
12            (SDS). Proteinase K (Boehinger Mannheim) was added to a  
13            concentration of 0.1 mg/ml and the cell extracts digested  
14            for 3 hours at 50°C. DNA was extracted 3 times with  
15            phenol and once with CHCl<sub>3</sub>. DNA was precipitated with 2  
16            volumes of ethanol, spooled and resuspended in 20 ml of  
17            10 mM Tris-HCl (pH 7.5), 1 mM EDTA. The solution was  
18            then made 10 µg/ml with (DNase free) RNase (Boehinger  
19            Mannheim) and incubated for 2 hr at 50°C. NaCl was added  
20            to 0.5 M and the solution extracted with phenol followed  
21            by CHCl<sub>3</sub>. DNA was precipitated with 2 volumes of  
22            ethanol, spooled, and resuspended in 10 mM Tris, 1 mM  
23            EDTA. The concentration was determined by routine  
24            spectrophotometric procedure at 260 nm wavelength.

1           2. From tissues:

2           Two grams original mass of primary tumor (designated  
3   MAC117 obtained from Memorial Sloan-Kettering Cancer  
4   Center Specimen code 31-26606) <sup>were</sup> ~~was~~ pulverized in a mortar  
5   and pestle at liquid nitrogen temperature, suspended in  
6   10 ml of 10 mM Tris-HCl (pH 7.5), 150 mM NaCl, 2 mM EDTA,  
7   reacted with proteinase K at 500 µg/ml (Boehinger  
8   Mannheim) and SDS at 0.5% at 37°C for 10 hr. The  
9   solution was then extracted twice with phenol and twice  
10   with the mixture of phenol:CHCl<sub>3</sub>: isoamyl alcohol at  
11   25:24:1 and once with CHCl<sub>3</sub>:isoamyl alcohol (24:1). DNA  
12   was precipitated by 2 volumes of ethanol removed by  
13   spooling, and resuspended in 1 mM Tris-HCl (pH 7.5), 0.2  
14   mM EDTA.

15           Electrophoretic analysis of DNA fragments using  
16           "Southern hybridization"

17           1. Restriction enzyme cleavage

18           Each sample of DNA (15 µg) was digested in 0.4 ml of  
19   100 mM Tris-HCl (pH 7.5), 50 mM NaCl, 5 mM MgCl<sub>2</sub>,  
20   100 µg/ml bovine serum albumin and 30 units of  
21   restriction enzyme (New England Biolabs) for 2 hr at  
22   37°C. Following reaction, 10 µg of tRNA was added and  
23   the solution extracted once with an equal volume of a  
24   mixture of phenol and CHCl<sub>3</sub> (1:1). Nucleic acids were

1 precipitated from the aqueous phase by addition of 2  
2 volumes of ethanol. Following centrifugation for 10 min  
3 at 12,000 x g (Eppendorf microfuge) the samples were  
4 washed once with 80% ethanol, dried to remove ethanol,  
5 and resuspended in 40  $\mu$ l distilled H<sub>2</sub>O.

6 2. Agarose gel electrophoresis

7 DNA samples were made 40 mM Tris acetate (pH  
8 7.2), 20 mM Na acetate, 1 mM EDTA, 5.0% glycerol, 0.05%  
9 bromophenol blue. Electrophoresis was conducted in a BRL  
10 H4 apparatus containing 400 ml 0.8% agarose, 40 mM Tris  
11 acetate (pH 7.2), 20 mM Na acetate, 1 mM EDTA and 1  $\mu$ g/ml  
12 ethidium bromide for about 16 hr at about 50 volts  
13 following conventional procedure. DNA was detected by  
14 irradiation with ultraviolet light.

15 3. Transfer to nitrocellulose

16 The agarose gel was treated twice for 15 min in 1  
17 liter of 0.5 M NaOH, 1.5 M NaCl, then twice for 30 min  
18 with 1 M NH<sub>4</sub>Ac, 20 mM NaOH. The agarose gel was then  
19 placed on a stack of filter paper saturated with 1 liter  
20 of 1 M NH<sub>4</sub>Ac, 20 mM NaOH. A sheet of nitrocellulose  
21 membrane (0.45  $\mu$ m pore size Schleicher & Schuell) was  
22 placed on top of the gel followed by dry filter paper.  
23 Transfer was allowed to occur overnight. DNA was fixed  
24 to <sup>+</sup><sub>1</sub> nitrocellulose by baking at 80°C in vacuo for 2 hr.

j

1     Hybridization to RNA and DNA blots

2             Hybridization was conducted in 20 ml of 40%  
3     formamide, 0.75 M NaCl, 0.075 M Na citrate, 0.05% BSA,  
4     0.05% polyvinyl pyrrolidone, 0.05% Ficoll 400 and 20 µg/ml  
5     sheared denatured calf thymus DNA. All hybridization was  
6     conducted for 16 hr at 42°C in a water bath. Following  
7     hybridization, nitrocellulose membranes were washed 2  
8     times for 20 min in 1 liter of 0.3M NaCl, 30 mM Na  
9     citrate, followed by washes in 15 mM NaCl, 1.5 mM Na  
10    citrate, first with and then without 0.1% sodium dodecyl  
11    sulfate. These final washes were at 42°C for v-erbB  
12    probes and at 52°C with pMAC117 and pE7 probes, vide  
13    infra. Autoradiography was conducted at -70°C with Kodak  
14    XAR5 film. Exposure times were 2 hr for Fig. ~~1A~~<sup>2A</sup> and 20  
15    min for Fig. ~~1AB~~<sup>2</sup>, 40 min for EGF receptor probe on Fig.  
16    4, and 4 hr for the pMAC117 probe of Fig. 4.

17    Generation of probes

18            A nucleic acid probe is defined as a fragment of DNA  
19    or RNA whose nucleotide sequence has at least partial  
20    identity with the sequence of the gene or its messenger  
21    RNA so as to enable detection or identification of the  
22    gene. Since a gene may have several fragments, there  
23    could be a plurality of probes for detecting the gene.

24            The probes used were the 0.5-kb Bam HI to Bam HI  
25    fragment combined with the 0.5-kb Bam HI to Eco RI

1 fragment of the v-erbB gene of AEV 11; the 1-kb BglI to  
2 Bam <sup>HI</sup> fragment of pMAC117; and the 2-kb Cla I fragment  
3 of pE7 as described by Xu, et al., (Nature 309:806, 1984).

#### 4 DNA

5 fragments were isolated by gel electrophoresis in 1% low  
6 melting point agarose gels (Bethesda Research Labs) in 40  
7 mM Tris acetate, 20 mM Na acetate, 1 mM EDTA, followed by  
8 melting of the gel at 70°C and extraction with phenol  
9 followed by CHCl<sub>3</sub> and ethanol precipitation. DNAs were  
10 made radioactive by using a nick-translation kit  
11 (Amersham) in which 50 µl reactions contained 250 µCi  
12 αP<sup>32</sup>dCTP (Amersham) and 0.5 µg DNA. Radioactive probe  
13 DNA was purified from unincorporated nucleotides by 2  
14 cycles of ethanol precipitation. Yields were above 2 x  
15 10<sup>8</sup> cpm/µg DNA. Before hybridization all probes were  
16 made single-stranded by treatment with 90% formamide.

#### 17 RNA electrophoresis and transfer to nitrocellulose

18 RNA samples (5 µg A431 polyadenylated RNA, obtained  
19 from National Institutes of Health, Bethesda, MD 21218)  
20 were treated for 5 min at 50°C in 50% formamide, 6.7%  
21 formaldehyde, 20 mM Mops (pH 7.0) (Sigma Biochemicals), 5  
22 mM Na acetate, 1 mM EDTA in 25 µl total volume.  
23 Electrophoresis was conducted in BRL H4 apparatus in  
24 250ml of 1.5% agarose, 20 mM Mops (pH 7.0), 5 mM Na

1 acetate, 1 mM EDTA, 1 µg/ml ethidium bromide at 40 volts  
2 for 16 hr. RNA was detected using ultraviolet light. The  
3 gel was soaked for 30 min at 20°C in 50 mM NaOH, followed  
4 by two 30 min washes in 1 M Tris (pH 7.5), followed by 30  
5 min in 3 M NaCl, 0.3 M Na citrate. Transfer to  
6 nitro-cellulose was accomplished by placing the gel atop  
7 a stack of filter paper saturated with 1.5 M NaCl, 0.15 M  
8 Na citrate, followed by 0.45 µM pore size nitrocellulose  
9 (Schleicher and Schuell), followed by dry filter paper.  
10 Transfer was allowed to proceed for 16 hr. The  
11 nitrocellulose filter was washed twice for 20 min in 0.3  
12 M NaCl, 30 mM Na citrate. RNA was fixed to the paper by  
13 baking at 80°C in vacuo for 2 hr.

14 DNA sequence analysis

15 DNA fragments containing the AccI-NcoI region  
16 (Fig. 1) were digested with either Nco I, Hinf I or Sau  
17 96I (New England Biolabs). These fragments were end-  
18 labeled in reactions of 50 µl containing 50 mM Tris-HCl  
19 (pH 7.2), 10 mM MgCl<sub>2</sub>, 0.1 mM dithiothreitol, 50 µg/ml  
20 BSA, 10 µCi <sup>α-<sup>32</sup>P</sup>-dNTP (Amersham--where x represents the  
21 correct nucleotide for fill-in), 2 units E. coli DNA  
22 polymerase large fragment (New England Biolabs).  
23 Following labeling, single-stranded material was prepared  
24 by electrophoresis. Samples were denatured in 30%

1 dimethyl sulfoxide, 1 mM EDTA and 0.05% bromophenol blue  
2 at 90°C for 2 hr. Samples were chilled and  
3 electrophoresed in acrylamide gels in a Bethesda Research  
4 Labs apparatus. DNA was detected by autoradiography and  
5 isolated by elution into 10 mM Tris-HCl (pH 7.0), 1 mM  
6 EDTA. Chemical degradation of DNA for sequence analysis  
7 was conducted using standard procedures. Cleavage at  
8 guanine (G) residues was conducted by reaction with  
9 dimethyl sulfonate at 22°C for 10 min. Cleavage at  
10 adenine (A) residues was conducted by 12 min reaction at  
11 90°C in 1.5 M NaOH, 1 mM EDTA. Cleavage at cytosine (C)  
12 residues was conducted using hydrazine in 2 M NaCl for 13  
13 min at 22°C. Cleavage at thymine (T) residues was  
14 conducted using hydrazine with no added NaCl for 10 min  
15 at 22°C. Following cleavage, all reactions were twice  
16 precipitated using ethanol and thoroughly dried. All  
17 samples were reacted with 1 M piperidine at 90°C for 30  
18 min. Piperidine was removed by evaporation in a Savant  
19 speed vac concentrator. Fragments were separated by  
20 electrophoresis in acrylamide gels (BRL HO apparatus) in  
21 8 M urea, 50 mM Tris-borate (pH 8.3), 1 mM EDTA.  
22 Detection of degraded ladder was by autoradiography using  
23 Kodak XAR5 film at -70°C.

1     Cloning of  $\lambda$ MAC117

2             High molecular weight DNA (6  $\mu$ g) from tumor MAC117  
3     (see above) was digested with 12 units restriction enzyme  
4     Eco RI (New England Biolabs) in a volume of 100  $\mu$ l for  
5     about one hour at 37°C. DNA was obtained by phenol  $\text{CHCl}_3$   
6     extraction and ethanol precipitation and resuspended in  
7     water at a concentration of 0.1  $\mu$ g/ml. This DNA (0.2  $\mu$ g)  
8     was ligated to  $\lambda$ ves  $\lambda$ B arms (Bethesda Research Labs)  
9     (1  $\mu$ g) using T4 DNA ligase (New England Biolabs) in a  
10    total volume of 20  $\mu$ l [50 mM Tris-HCl pH 7.4, 10 mM  $\text{MgCl}_2$   
11    10 mM dithiothreitol, 0.5 mM spermidine, 1 mM ATP]. This  
12    mixture of ligated DNAs was packaged into infectious  
13    bacteriophage particles using the Packagene system  
14    (Promega Biotec). These particles were used to infect  
15    bacteria BNN45 and about  $8 \times 10^5$  individual phage plaques  
16    were obtained.

17            These phage plaques were screened for individual  
18    plaques containing DNA homologous to the v-erbB probes  
19    (described above) using standard procedures. Briefly,  
20    bacterial culture <sup>+</sup>plaques containing approximately 15,000  
21    plaques were grown overnight. Sterile nitrocellulose  
22    discs (Scheicher and Schuell) were applied to the dish,  
23    removed and allowed to air dry for about 90 minutes. The



1 discs were then treated with 0.2 M NaOH, 1.5 M NaCl  
2 followed by 0.4 M Tris-HCl pH 7.5 followed by 0.3 M NaCl  
3 0.03 M Na citrate and baked in vacuo for two hours at  
4 80°C. These discs were then exposed to hybridization and  
5 washing conditions identical to those described for Fig.  
6 <sup>2</sup> using the identical v-erbB probe. Washing conditions  
7 <sup>1</sup> were also identical to those for Fig. <sup>2</sup> 1. Hybridization  
8 was detected by autoradiography at -70°C for 16 hours.  
9 Single hybridizing phage plaques were obtained by three  
10 successive hybridization experiments (as described above)  
11 to isolate a pure phage culture.

12 ~~To clone the 6-kbp fragment,~~ DNA from MAC117 was  
13 digested with Eco RI, then ligated into bacteriophage  
14  $\lambda$ gWES, packaged in vitro, and transferred to Escherichia  
15 coli (E. coli) strain BNN45 by infection following  
16 standard techniques well known in the art. A library of  
17  $4 \times 10^5$  bacteriophages was screened by plaque  
18 hybridization with radioactive v-erbB DNA. Ten of 14  
19 hybridizing phages contained a 6-kbp Eco RI fragment.  
20 Figure <sup>1</sup> shows the physical map of one of these phages,  
21  $\lambda$ MAC117, and pMAC117, a PUC12 subclone containing a 2-kbp  
22 Bam HI fragment of  $\lambda$ MAC117 that hybridized with v-erbB  
23 probes. The region of pMAC 117 to which v-erbB  
24 hybridized most intensely was flanked by Acc I and Nco I  
25 sites. Human repetitive sequences were also localized  
26 (Fig. <sup>1</sup> 2, region demarcated by arrows).

1        A deposit of pMAC117 cloned in E. coli has been made  
2        at the American Type Culture Collection (ATCC), Bethesda,  
3        Md. under accession number 53408. Upon issuance of a  
4        patent, the culture will continue to be maintained for at  
5        least 30 years and made available to the public without  
6        restriction subject, of course, to the provisions of the  
7        law in this respect.

8        As shown in Fig. 1A, DNA prepared from tissue of a  
9        human mammary carcinoma, MAC117, showed a pattern of  
10       hybridization that differed both from that observed with  
11       DNA of normal human placenta and from that observed with  
12       the A431 ~~squamous~~<sup>epithelial</sup>-cell carcinoma line, which contains  
13       amplified epidermal growth factor (EGF) receptor genes.  
14       In A431 DNA, four Eco RI fragments were detected that had  
15       increased signal intensities compared to those of  
16       corresponding fragments in placenta DNA (Fig. 1A). In  
17       contrast, MAC117 DNA contained a single 6-kilobase pair  
18       (kbp) fragment, which appeared to be amplified compared  
19       to corresponding fragments observed in both A431 and  
20       placenta DNA's (Fig. 1A). These findings indicate that  
21       the MAC117 tumor contained an amplified DNA sequence  
22       related to, but distinct from, the cellular erbB  
23       proto-oncogene.

1 By digestion of pMAC117 with Bgl I and Bam HI, it  
2 was possible to generate a single-copy probe homologous  
3 to v-erbB. This probe detected a 6-kb Eco RI fragment  
4 that was amplified in MAC117 DNA and apparently increased  
5 in A431 cellular DNA relative to normal DNA (Fig. <sup>2</sup>/<sub>1</sub>B).  
6 The sizes of the fragment corresponded to the amplified  
7 6-kb Eco RI fragment detected in MAC117 DNA by means of  
8 v-erbB (Fig. <sup>2</sup>/<sub>1</sub>A). Hybridization to Southern blots  
9 containing serial dilutions of MAC117 genomic DNA  
10 indicated an approximate amplification of 5- to 10-fold  
11 when compared to human placenta DNA.

12 The nucleotide sequence of the portion of pMAC117  
13 located between the Nco I and Acc I sites contained two  
14 regions of nucleotide sequence homologous to v-erbB  
15 separated by 122 nucleotides (Fig. <sup>1</sup>/<sub>1</sub>). These regions  
16 shared 69 percent nucleotide sequence identity with both  
17 the v-erbB and the human EGF receptor gene. The  
18 predicted amino acid sequence of these regions was 85  
19 percent homologous to two regions that are contiguous in  
20 the EGF receptor sequence. Furthermore, these two  
21 putative coding regions of the MAC117 sequence were each  
22 flanked by the AG and GT dinucleotides that border the  
23 exons of eukaryotic genes. These findings suggest that  
24 the sequence shown in Fig. <sup>2</sup>/<sub>1</sub> represents two exons,  
25 separated by an intron of a gene related to the erbB/EGF  
26 receptor gene.

1           The predicted amino acid sequence of the  $\lambda$ MAC117  
2     putative exons is homologous to the corresponding  
3     sequences of several members of the tyrosine kinase  
4     family. The most striking homology was observed with the  
5     human EGF receptor or erbB (Fig. 3). In addition, 42  
6     percent to 52 percent homology with the predicted amino  
7     acid sequences of other tyrosine kinase-encoding genes  
8     was observed. At 25 percent of the positions there was  
9     identity among all the sequences analyzed (Fig. 3). A  
10    tyrosine residue in the  $\lambda$ MAC117 putative coding sequence,  
11    conserved among the tyrosine kinases analyzed, is the  
12    site of autophosphorylation of the src protein (Smart et  
13    al., Proc. Natl. Acad. Sci. USA. 78:6013, 1981).

14           The availability of cloned probes of the MAC117 gene  
15    made it possible to investigate its <sup>the</sup> expression in a  
16    variety of cell types. The MAC117 probe detected a  
17    single 5-kb transcript in A431 cells (Fig. 4). Under the  
18    stringent conditions of hybridization utilized, this  
19    probe did not detect any of the three RNA species  
20    recognized by EGF receptor complementary DNA. Thus,  
21    MAC117 represents a new functional gene within the  
22    tyrosine kinase family, closely related to, but distinct  
23    from the gene encoding the EGF receptor.

1           There is precedent for the identification of genes  
2   related to known oncogenes on the basis of their  
3   amplification in human tumors. For example, the high  
4   degree of amplification of N-myc in certain malignancies  
5   made it detectable by means of the myc gene as a  
6   molecular probe (Schwab, Nature 305:245, 1983; Kohl et  
7   al., Cell 35:349, 1983). In the present study, a five-  
8   to tenfold amplification of a v-erbB-related gene in the  
9   MAC117 mammary carcinoma made it possible to identify  
10   this sequence against a complex pattern of EFG receptor  
11   gene fragments.

12  
13  
14           The MAC117 coding sequence, as determined by  
15   nucleotide and predicted amino acid sequence, is most  
16   closely related to the erbB/EGF receptor among known  
17   members of the tyrosine kinase family. The two genes are  
18   distinct, however, as evidenced by the sequence diversity  
19   and transcript size. Detailed structural analysis of the  
20   complete coding sequence would further elucidate the role  
21   and function of this v-erbB-related gene.

To this purpose we have isolated cDNAs with a complexity of over 4.5 kb from the MAC117 mRNA (Kraus et al., 1987). <sup>EMBO Journal 6:605-610,</sup> A restriction map is shown in Fig. 5A. An oligo (dT) primed normal human fibroblast cDNA library (Okayama and Berg, 1983) was screened with a 0.8 kbp Acc I DNA fragment from <sup>the 3' terminus of</sup> a genomic clone of MAC117 (Fig. 1). The largest plasmid obtained, pMAC137, carried a 2-kbp insert comprising 1.5 kbp of 3' coding information and 3' untranslated sequence. The remaining coding information upstream was obtained from three phage clones,  $\lambda$ MAC30,  $\lambda$ MAC10' and  $\lambda$ MAC14-1, identified in a randomly primed MCF-7 cDNA library (Walter et al., 1985; Fig. 5A).

Fig. 5B illustrates 3 probes, a, b, and c, representing the 5' end, a middle portion, and the entire coding region, respectively, which were employed in subsequent studies elucidating the role and function of this v-erbB-related gene.

To assess the role of MAC117 in human mammary neoplasia, we compared mRNAs of 16 mammary tumor cell lines to normal human fibroblasts, M413, and a human mammary epithelial cell line, HBL100. Increased expression of an apparently normal size 5-kb transcript was detected in 8 of 16 tumor cell lines when total cellular RNA was subjected to Northern blot analysis. An aberrantly sized erbB-2 mRNA was not detected in any of the cell lines analyzed (Kraus et al., 1987). <sup>EMBO Journal 6:605-610,</sup> To quantitate more precisely the amount of MAC117 transcript in eight mammary tumor cell lines which overexpress MAC117, serial 2-fold dilutions of total cellular RNA were subjected to dot blot analysis using human  $\beta$  actin as a control for the amount of RNA applied to the nitrocellulose filters. The highest levels of MAC117 mRNA, which ranged from 64- to 128-fold over that

of our controls, were observed in the cell lines MDA-MB453, SK-BR-3, MDA-MB361, and BT474. Moreover, MAC117 mRNA levels were increased 4- to 8-fold in four cell lines including BT483, MDA-MB175, ZR-75-30, and ZR-75-1 (Fig. 6).

To determine if the overexpression of MAC117 mRNA resulted in a steady state increase of its encoded gene product, we developed a specific immunoblot assay. Antisera were raised against a synthetic peptide whose sequence corresponded to a portion of the putative tyrosine kinase domain of MAC117. As this region is partially conserved between the encoded proteins of the EGFR and MAC117 genes, we tested its specificity using A431 and SK-BR-3 cell lines which overexpress EGFR or MAC117 mRNA, respectively. As shown in Fig. 7A, a specific band of ~ 185 kd was detected in extracts of SK-BR-3 but not in A431 cells. This band was not detected when the antibody was preincubated with the synthetic peptide corresponding to its antigen. To estimate the relative amounts of MAC117 protein in different mammary tumor cell lines, immunoblot analysis was conducted using equivalent amounts of total cellular protein. As shown in Fig. 7B, an intense band of protein was detected in extracts of SK-BR-3 and a less intense but readily detectable band in extracts of ZR-75-1. No MAC117 protein was detected in extracts of MCF-7, a mammary tumor cell line, that did not display overexpression of erbB-2 mRNA. We interpret these results to indicate that substantially more erbB-2 protein is found in both SK-BR-3 and ZR-75-1 than in MCF-7 cells where the amount of protein escapes the sensitivity of the assay. These observations indicated that elevated mRNA levels of MAC117 are translated into MAC117 proteins. This demonstrated that gene amplification of MAC117 results in overexpression of mRNA and protein of MAC117 in human mammary tumor cells. Furthermore, increased mRNA and protein levels are observed in mammary tumor cells in the absence of gene amplification suggestive

for an additional mechanism as a result of which mRNA and protein of our novel v-erbB-related gene can be found overexpressed in tumor cells (Kraus et al., 1987).

To directly assess the effects of MAC117 overexpression on cell growth properties, we assembled a full length normal human MAC117 clone from overlapping cDNA clones (<sup>Figs. 5A,B</sup>~~(Fig. 5)~~) linked to the transcriptional initiation sequences of either the Moloney murine leukemia virus long terminal repeat (MuLV LTR) or the SV40 early promoter in expression vectors in order to express a normal coding sequence of MAC117 in NIH3T3 cells (Fig. 9) (DiFiore et al., <sup>Science 237:178-182</sup>1987).

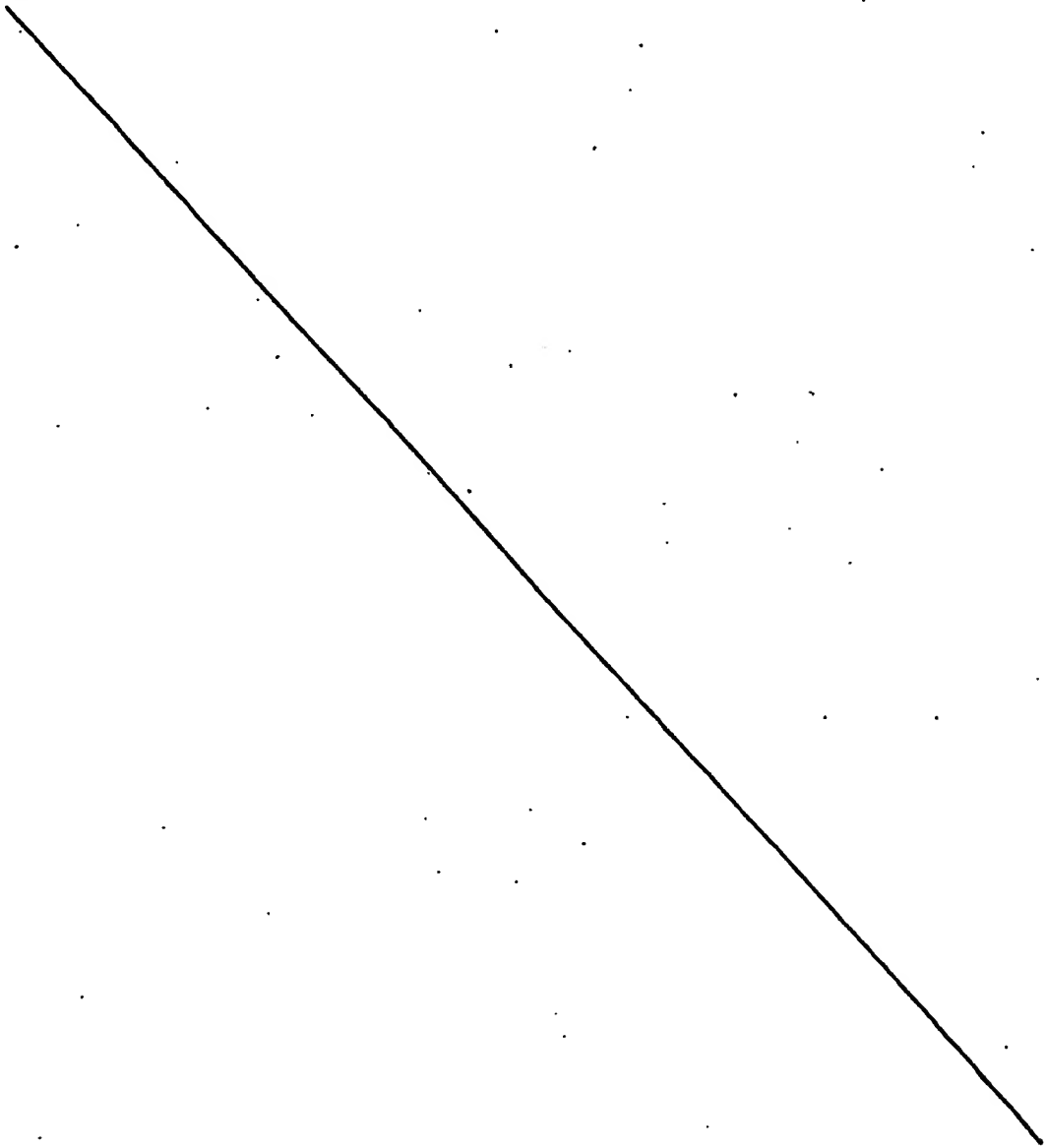
Previous studies have indicated different strengths of LTR and the SV40 promoters in these cells (Gorman et al., Proc. Natl. Acad. Sci. USA 79, 6777, 1982). Because of the presence of the MuLV donor splice site close to the 5' LTR (Shinnick et al., Nature 293, 543, 1981), we engineered one of the LTR-based vectors (LTR-1/MAC117) to contain an acceptor splice site immediately upstream of the translation initiation codon of the MAC117 coding sequence (Fig. 9). This vector was constructed in order to ensure correct splicing of the message even if a cryptic splice acceptor site were present within the MAC117 open reading frame. In the SV40-based expression vector (SV40/MAC117) the erbB-2 coding sequence replaced the neomycin-resistance gene of pSV2/neo (Southern et al., J. Mol. Appl. Genet. 1, 327, 1982) (Fig. 9). To assess the biologic activity of our human MAC117 vectors, we transfected NIH/3T3 cells with serial dilutions of each DNA. As shown in Table <sup>1</sup>~~1~~, LTR-1/MAC117 DNAs induced transformed foci at high efficiency of  $4.1 \times 10^4$  focus-forming units per picomole of DNA (ffu/pM). In striking contrast, the SV40/erbB-2 construct failed to induce any detectable morphological alteration of NIH/3T3 cells transfected under identical assay conditions (Table <sup>1</sup>~~1~~). Since the SV40/erbB-2



construct lacked transforming activity, these results demonstrated that the higher levels of MAC117 expression under LTR influence correlated with its ability to exert transforming activity. To compare the growth properties of NIH/3T3 cells transfected by these genes, we analyzed the transfectants for anchorage-independent growth in culture, a property of many transformed cells. The colony-forming efficiency of a LTR-1/MAC117 transformant was very high and comparable to that of cells transformed by LTR-driven v-H-ras and v-erbB (Table 1). Moreover, the LTR-1/MAC117 transfectants were as malignant in vivo as cells transformed by the highly potent v-H-ras oncogene and 50-fold more tumorigenic than cells transfected with v-erbB. In contrast, SV40/MAC117 transfectants lacked anchorage-independent growth in vitro and did not grow as tumors in nude mice even when  $10^6$  cells were injected (Table 1).

To compare the level of overexpression of the 185-kd protein encoded by MAC117 in human mammary tumor cell lines possessing amplified MAC117 genes with that of NIH/3T3 cells experimentally transformed by the MAC117 coding sequence, we compared MAC117 specific protein amounts by Western blotting (DiFiore et al., *Science* 237: 178-182, 1987). An anti-MAC117 peptide serum detected several discrete protein species ranging in size from 150 to 185 kd in extracts of MDA-MB361 and SK-BR-3 mammary tumor cell lines, as well as LTR/MAC117 NIH/3T3 transformants (Fig. 10). The relative levels of the 185-kd MAC117 product were similar in each of the cell lines and markedly elevated over that expressed by MCF-7 cells, where the 185-kd protein was not detectable under these assay conditions (Fig. 10). Thus, human mammary tumor cells which overexpressed the MAC117 gene demonstrated levels of the MAC117 gene product capable of inducing malignant transformation in a model system.

Overexpression of proto-oncogenes can cause cell transformation in culture and may function in the development of human tumors. Amplification of a normal



1     ras gene or its increased expression under the control of,  
2     a retroviral long terminal repeat (LTR) induces  
3     transformation of NIH 3T3 cells (Chang et al., Nature  
4     297:479, 1982). Expression of the normal human  
5     sis/PDGF-2 coding sequence in NIH 3T3 cells, which do not  
6     normally express their endogenous sis proto-oncogene,  
7     also leads to transformation (Gazit et al., Cell 39:89,  
8     1984; Clarke et al., Nature 308:464, 1984). In Burkitt  
9     lymphoma, a chromosomal translocation involving myc  
10    places its normal coding sequence under the control of an  
11    immunoglobulin gene regulatory sequence. The resulting  
12    alteration in myc expression is likely to be causally  
13    related to tumor development (Nishikura et al., Science  
14    224:399, 1984). The observation of amplification of myc  
15    or N-myc in more malignant phenotypes of certain tumors  
16    has supported the idea that overexpression of these genes  
17    can contribute to the progression of such tumors. The  
18    erbB/EGF receptor gene is amplified or overexpressed in  
19    certain tumors or tumor cell lines. The five- to tenfold  
20    amplification of the v-erbB-related gene of the present  
21    invention in a mammary carcinoma indicates that increased  
22    expression of this gene may have provided a selective  
23    advantage to this tumor. The isolation of a new member  
24    of the tyrosine kinase gene family amplified in a human  
25    mammary carcinoma in accordance with the present  
26    invention, makes possible the elucidation of the role of  
27    this gene in human malignancy.

1     Use of Specific Nucleic Acid Probes

j     2             As demonstrated in Figures <sup>2</sup>1 and 4, the isolation  
3     and use of a Bgl I to Bam HI restriction fragment of  
4     pMAC117 to specifically detect the gene and its mRNA  
5     product has been set forth. The importance of this  
6     technique, involving this probe and others like it, is  
7     that the biological functions of the gene described here  
8     can be determined and these functions related to  
9     practical application, some of which are listed below.

- 10    1. Isolation of cloned cDNA. This involves the use of  
11       probes specific for the gene described herein; an  
12       example is the Bgl I-Bam HI fragment of pMAC117.  
13       These probes are made radioactive by standard  
14       techniques, such as those noted above, and screening  
15       of the libraries of cDNA clones is done using  
16       standard methods analogous to those described in  
17       "Cloning of \MAC117" above. This approach was employed  
      to clone cDNA comprising the entire coding region of  
      this gene, the restriction map of which is shown in  
      Fig. 5A.
2. Use of cDNA clones. Due to the fact that cDNA clones  
      contain complete information for encoding the  
      protein, these cDNA clones provide a "second  
      generation" of specific probes for the gene described  
      herein. Such probes are shown in Fig. 5B. Their appli-  
      cation for hybridization analysis is demonstrated in Fig. 6  
      and Fig. 8. As shown in Fig. 8, the availability of  
j     probes, such as probe <sup>6</sup>1 in Fig. 5B, facilitates the com-  
      prehensive hybridization analysis of the entire coding

region of this gene or any defined part of it. In addition, the complete coding information allows the expression of the protein product in a heterologous system. Such systems utilize strong and/or regulated transcription promoters placed in such a way as to direct overexpression of the gene. Techniques for accomplishing expression of the gene are well known in the art and can be found in such publications as Rosenberg et al., Methods in Enzym. 101, 123 (1983); Guarante, L., Methods in Enzym. 101, 181 (1983). The coding region of our novel v-erbB-related gene was overexpressed under the transcriptional control of MuLV-LTR or SV40 early promoter. Thereby, high expression levels were achieved with MuLV-LTR which caused the neoplastic transformation of transfected cells. These cells can be used as a source to rescue infectious recombinant virus which might prove useful to infect heterologous cells not susceptible to DNA transfection. In addition, these cells serve as a source for high and defined levels of antigen for this novel v-erbB-related gene.

3. Preparation of antibodies specific for the protein product of the gene. Of course, the identification and knowledge of the gene allows its product, protein, for example, to be detected. Poly- or monoclonal antibodies are prepared against said

protein by standard techniques, often by commercially available services. The critical reagent in the production of antibodies is the antigen (protein) used. In this case, the antigens are either the peptides chemically synthesized by standard and commercially available techniques according to the predicted amino acid sequences derived from the nucleic acid sequence of the gene or its corresponding cDNA. Another potential antigen is the protein itself encoded by the gene and purified from the heterologous expression systems as described above. The antibodies are then employed by standard immunological techniques for the specific detection or diagnostic purposes. Such antibodies were raised against a peptide representing amino acids 35 through 49 of the peptide ~~sequence of claim 4~~. The specificity of these antibodies in detecting the gene product of this ~~novel v-erbB-related gene~~ <sup>also j<sup>2</sup></sup> ~~is~~ <sup>also j<sup>2</sup></sup> demonstrated in Fig. 7A. As shown in Fig. 7B and Fig. 10, these antibodies can be utilized to detect the over-expression of the protein product of our novel v-erb-B-related gene in human mammary tumor cells.

#### Further Applications of the Gene:

Having the knowledge of the gene allows preparing specific nucleic acid probes to detect the gene described here or its mRNA product. The probes are, of course,

derived from the gene, such as the Bgl I-Bam HI fragment of pMAC117 used in Figures <sup>2</sup>1 and 4, or alternatively such probes are derived from <sup>1</sup>other regions of the gene or its corresponding cDNA corresponding cDNA, as shown in Fig. <sup>3</sup>5. The use of nucleic acid probes in the molecular diagnosis of human cancer has been documented (Taub et al., Proc. Natl. Acad. Sci. USA 79, 7837 (1983); Schwab et al., Proc. Natl. Acad. Sci. USA 81, 4940 (1984)). The finding that the gene described here is amplified in a human mammary carcinoma indicates that alterations occur to this gene in human disease. Thus, detection of the amplification <sup>or increase expression</sup> of this gene provides useful diagnostic tools for the detection and treatment of human mammary carcinoma or other malignancies resulting from the ~~v-erbB~~ related gene. Hence, diagnostic kits which contain as their principal component specific nucleic acid probes for this gene <sup>or its mRNA transcript</sup> are of commercial value. The probe is used in analyses similar in concept to those shown in Figure <sup>1</sup>1 and Figure 4 for the detection of gene amplification structure or the expression of mRNA.

Specific antibody reagents (as described above) capable of detecting the protein product of the gene described herein are employed in a way similar to the use of specific nucleic acid probes. In other words, the expression of aberrant forms and amounts of a gene product is a measure of the related neoplastic condition

(Nishikura et al., Science 224, 399 (1984); Srivastava, et al., Proc. Natl. Acad. Sci. USA 82, 38-42 (1985)).

The detection of the aberrant expression of the protein product of the gene is of importance in the diagnosis of human cancers. As shown in Fig. 7 and Fig. 10, antibodies generated against peptides derived <sup>from</sup> ~~from~~ parts of the amino acid sequence of ~~claim 4~~ <sup>claim 1</sup> specifically detect the protein product of the gene described in ~~claim 1~~ in human tumor cells. Antibody reagent (produced as described above) is, of course, the critical reagent of the diagnostic kits for this purpose. Such antibody reagents are then employed in such standard methodologies as immunoprecipitation, western blot analysis, immunofluorescence analysis and the like well known in the art. The determination of amplification in a human mammary carcinoma of the gene described here indicates that overexpression (or other abnormality) of the protein product of this gene is functionally important, thus diagnostically relevant. This relevance is further substantiated by the <sup>ob</sup> ~~ob~~servations that gene amplification of this gene is associated with overexpression of its mRNA and protein in human mammary tumor cells and that protein levels observed in human mammary tumor cell lines exhibiting gene amplification of this gene are sufficient to induce neoplastic transformation of NIH/3T3 cells in vitro. Furthermore, a recent report (Slamon et al., 1987) correlates gene amplification of this novel erbB-related gene with a reduced disease free survival in breast cancer patients, suggesting the potential usefulness of analysis of this gene for its gene product as a diagnostic parameter in the clinical <sup>setting</sup>.



A diagnostic test in accordance with the present invention involves, for example, material obtained by surgical biopsy of potential tumor material. Such material is then analyzed by one or more procedures as follows.

1. DNA is isolated from the sample by standard methods (see above). The DNA is then analyzed by established methods, such as Southern blot hybridization using standard techniques similar to those used in the analysis shown in Figure 2. Gene-specific probes (described above) are made radioactive by standard techniques and used for detecting genetic abnormalities. Such abnormalities include gene amplification, as seen in the MAC117 tumor sample and tumor cell lines in Fig. 8. or gene rearrangement, as detected by aberrantly migrating bands of hybridization.
2. RNA is isolated from the tumor sample by standard methods (see above). This RNA is analyzed by blot hybridization techniques similar to those described in Figure 4. Gene-specific probes (described above) are made radioactive by standard techniques and used for detecting the mRNA products of the erbB-related gene described here. Such abnormalities include overexpression or abnormal forms of RNA. Overexpression of an apparently normal sized mRNA is shown in 8 human mammary tumor cell lines in Fig. 6. In addition, mRNA amount may also be quantitated by spot hybridization

procedures in which serial dilutions of RNA <sup>are</sup> ~~is~~ fixed to nitrocellulose filter and the mRNA of v-erb-B-related gene described here detected by hybridization. Such a procedure has been employed in Fig. 6B. The foregoing techniques are standard. This allows detection of mRNA overexpression or alteration of structure.

When antigens or proteins (polypeptides) are to be analyzed, the proteins are separated according to molecular size, for example by gel electrophoresis, transferred to nitrocellulose membranes and the protein product of the erbB-related gene described here detected by reaction with specific antibodies, described above. Such a test is able to detect alterations in the quantity of protein as well as abnormal protein forms. With such an approach protein levels of the v-erb-B-related gene have been detected in human mammary tumor cell lines (Fig. 7, Fig. <sup>10</sup>~~11~~). <sup>1</sup>

In addition, specific antibodies may be used in the analysis of histological sections. These techniques, which are well known for other antibody specificities, involve the thin sectioning of biopsied material from a potential tumor, followed by reaction with specific antibodies. The antibody-antigen reaction is then made visible by a variety of standard methods including labeling with fluorescently tagged or ferritin tagged

1 second antisera and the like. Such detection systems  
2 allow the detection of the localized aberrant display of  
3 the protein product of the erbB-related gene described  
4 here.

5 In addition, although the <sup>R</sup> demonstrated genetic  
6 abnormality (shown in Figure 1) of the gene described  
7 here occurs in human mammary carcinoma, genetic  
8 abnormalities may also be associated with other  
9 clinically important syndromes of neoplastic or other origin.  
10 Genetic abnormalities have long been known to be involved  
11 in thalassemias, for example.

12 Knowledge of the erbB-related gene described here  
13 also makes possible a means of cancer treatment. If it  
14 is found that some cancers display abnormally high  
15 quantities of the gene product on their surface, such  
16 tumors can be treated with antibodies specific for the  
17 gene product which <sup>have</sup> ~~has~~ been conjugated to a toxic  
18 substance, such as radioactive markers, biological  
19 modifiers or toxins and the like. Another treatment  
20 modality involves a similar assumption of  
21 overexpression. In this approach, a specific natural  
22 product, even if unidentified but which has high binding  
23 affinity for the protein product of the gene described

1 here is used to target toxins to the tumor cells. This  
2 treatment modality is supported by the finding, reported  
3 here, of distinct but limited homology of this gene  
4 product to the EGF receptor. If a ligand analogous to  
5 EGF exists for the erbB-related gene described here, it  
6 may serve as such a targeting agent.

7 Diagnostic kits for the detection of the protein  
8 product of the erbB-related gene. Kits useful for the  
9 diagnosis of human cancers having abnormalities of this  
10 gene are now disclosed.

11 a) Kits designed to detect the protein by immunoblotting  
12 These kits preferably comprise containers containing  
13 (a) homogenization solution (50 mM Tris-HCl pH 7.5,  
14 1% sodium dodecyl sulfate and 0.1%  $\beta$ -mercaptoethanol)  
15 for the extraction of protein sample from biopsied  
16 material from putative tumors; (b) reagents for the  
17 preparation of immunoblots of the protein samples  
18 (acrylamide gels are prepoured and contain 7.5%  
19 acrylamide, .025% bis acrylamide, 0.38 M Tris-HCl pH  
20 8.8, 0.1% sodium dodecyl sulfate; the nitrocellulose  
21 sheets will be formed to the gel size; and transfer  
22 buffer 0.25 M Tris-glycine pH 8.8, 30% methanol);  
23 specific antibody reagents for the detection of the

1 protein product of the erbB-related gene (antisera  
2 directed against the protein product of erbB- related  
3 gene described here and reaction buffer containing  
4 0.1 M Tris-HCl pH 7.5, 5.0 M EDTA, 0.25% gelatin,  
5 0.1% nonidet P-40); and reagents and instructions for  
6 the visualization and interpretation of  
7 antibody-antigen interaction (these include  
8 radioactive protein A; biotin conjugated second  
9 antiserum, or peroxidase conjugated second  
10 antiserum). While this kit includes components  
11 ordinarily found and well known in the art, the  
12 critical component is the gene product-specific  
13 antibodies and buffers or media for performing  
14 immunological tests. The antibodies are derived or  
15 prepared as described above from either the peptide  
16 sequence predicted from the nucleotide sequence of  
17 the gene or its mRNA or from the protein product  
18 itself through standard immunization procedures.

19 b) Kits designed for the detection of the protein  
20 product of the erbB-related gene in tissue sections.  
21 Such kits include instructions for preparation of  
22 sections; instructions and standard reagents for the  
23 preparation of slides for microscopy;  $H_2O_2$  for  
24 removal of endogenous peroxidase; instructions for  
25 incubation with antibodies specific for the protein

1 product of the erbB-related gene described here in a  
2 buffer solution preferably containing phosphate  
3 buffered saline; and second antibodies for detection  
4 (these may be coupled to peroxidase, biotin, or  
5 ferritin); and instructions for visualization of  
6 detection complex. In addition the kits may include:  
7 reagents and instructions<sup>tions</sup> for the preparation of  
8 sections from biopsied putative tumor material;  
9 specific antibody reagents for the protein product of  
10 erbB-related gene described here and instructions for  
11 its reaction with the tissue section; and reagents  
12 and instructions for the detection of the  
13 protein-antibody interaction either by  
14 immunofluorescence, ferritin conjugated second  
15 antibodies or other standard methods well known in  
16 the art.

17 A Method for the Treatment of Human Cancers which Express  
18 High Levels of the Protein Product of the Gene Described  
19 herein.

20 This method involves administering to the patient  
21 one of two types of reagent which preferentially binds  
22 cells expressing high levels of the protein product of  
23 the erbB-related gene described here. These reagents are  
24 either antibodies directed against the protein product or

1 a ligand, which is likely to exist because of the  
2 homology of the gene to a growth factor receptor. The  
3 ligand is isolated by standard techniques using the  
4 intrinsic protein kinase activity of the protein product  
5 of the erbB-related gene. Extracts of body fluids and  
6 cell culture supernatants are incubated with the protein  
7 and  $\gamma$ - $^{32}\text{P}$ ATP. The presence of ligand is inferred by  
8 incorporation of  $^{32}\text{P}$  into the protein. The ligand is  
9 then purified by standard techniques such as ion exchange  
10 chromatography, gel permeation chromatography,  
11 isoelectric focusing, gel electrophoresis and the like.  
12 The natural ligand or antibody is tagged with one or more  
13 agents which will cause injury to cells to which they  
14 bind. Such tagging systems include incorporation of  
15 radioactive or biological toxins. The present discovery  
16 of amplification of the erbB-related gene makes it likely  
17 that some tumors carry large amounts of the corresponding  
18 protein. Hence, the two type-specific agents will bind  
19 in larger amounts to the protein present in the body and  
20 thus direct the toxic effects of the reagents to these  
21 cells.

22 It is understood that the examples and embodiments  
23 described herein are for illustrative purposes only and  
24 that various modifications or changes in light thereof  
25 will be suggested to persons skilled in the art and are  
26 to be included within the spirit and purview of this  
27 application and the scope of the appended claims.

1     a ligand, which is likely to exist because of the  
2     homology of the gene to a growth factor receptor. The  
3     ligand is isolated by standard techniques using the  
4     intrinsic protein kinase activity of the protein product  
5     of the erbB-related gene. Extracts of body fluids and  
6     cell culture supernatants are incubated with the protein  
7     and  $\gamma$ -p<sup>32</sup> ATP. The presence of ligand is inferred by  
8     incorporation of p<sup>32</sup> into the protein. The ligand is  
9     then purified by standard techniques such as ion exchange  
10    chromatography, gel permeation chromatography,  
11    isoelectric focusing, gel electrophoresis and the like.  
12    The natural ligand or antibody is tagged with one or more  
13    agents which will cause injury to cells to which they  
14    bind. Such tagging systems include incorporation of  
15    radioactive or biological toxins. The present discovery  
16    of amplification of the erbB-related gene makes it likely  
17    that some tumors carry large amounts of the corresponding  
18    protein. Hence, the two type-specific agents will bind  
19    in larger amounts to the protein present in the body and  
20    thus direct the toxic effects of the reagents to these  
21    cells.

22           It is understood that the examples and embodiments  
23    described herein are for illustrative purposes only and  
24    that various modifications or changes in light thereof  
25    will be suggested to persons skilled in the art and are  
26    to be included within the spirit and purview of this  
27    application and the scope of the appended claims.



1     CLAIMS

2           1. An isolated cloned human gene having at least in  
3     part the following nucleotide sequence:

4     GTCTACATGGGTGCTTCCCATTCAGGGGATGAGCTACCTGGAGGATGTGCGGCTCG  
5     TACACAGGGACTTGGCCGCTCGGAACGTGCTGGTCAAGAGTCCCAACCATGTCAAAA  
6     TTACAGACTTCGGGCTGGCTCGGCTGCTGGACATTGACGAGACAGAGTACCATGCAG  
7     ATGGGGGCAAGGTTAGGTGAAGGACCAAGGAGCAGAGGAGGCTGGGTGGAGTGGTGTG  
8     TAGCCCATGGGAGAACTCTGAGTGGCCACCTCCCCACAACACACAGTTGGAGGACTT  
9     CCTCTTCTGCCCTCCCAGGTGCCCATCAAGTGGATGGCGCTGGAGTCCATTCTCCGC  
10    CGGCGGTTACCCACCAGAGTGATGTGTGGAGTTATGGTGTGTGATGGGGGGTGTG  
11    GGAGGGGTGGGTGAGGAGCCATGG---

12    wherein A, T, C and G represent adenine, thymine,  
13    cytosine and guanine nucleotides, respectively.

14           2. The gene of Claim 1 having the characteristics  
15    of ATCC deposit number 53408.

16           3. Nucleic acid probes having specific binding  
17    affinity for at least a part of the gene of Claim 1 or  
18    for a nucleic acid derivative thereof.

1           4. A polypeptide having at least in part the  
2           following amino acid sequence:

3           GlyMetSerTyrLeuGluAspValArgLeuValHisArgAspLeuAlaAlaArgAsn  
4           ValLeuValLysSerProAsnHisValLysIleThrAspPheGlyLeuAlaArgLeu  
5           LeuAspIleAspGluThrGluTyrHisAlaAspGlyGlyLysValProIleLysTrp  
6           MetAlaLeuGluSerIleLeuArgArgArgPheThrHisGlnSerAspValTrpSer  
7           TyrGly---.

8           5. Antibody having specific binding affinity for at  
9           least a portion of the polypeptide of Claim 4.

10          6. A test kit for detecting genetic abnormalities  
11          related to the gene of Claim 1 in humans comprising  
12          containers containing specific nucleic acid probes of  
13          Claim 3 and instructions for performing test with said  
14          probes.

15          7. Antibody reagent kit for detecting the  
16          polypeptide of Claim 4 comprising containers containing  
17          antibodies of Claim 5, immunological reagents and  
18          instructions for using the kit.

19          8. A method of diagnosing human cancer related to  
20          the gene of Claim 1 comprising:

1           (a) detecting amplification rearrangement or  
2 over-expression of the gene of Claim 1 by hybridizing  
3 nucleic acid derived from a tissue sample of a human  
4 suspected of said cancer with the nucleic acid probes of  
5 Claim 3; or

6           (b) detecting abnormal expression of the protein  
7 product of the gene of Claim 1 by reacting a body sample  
8 of a human suspected of said cancer with antibodies of  
9 Claim 5.

10           9. A method of inhibiting malignancy caused by  
11 erb-B related gene comprising reacting protein produced  
12 by said gene with antibody having specific binding  
13 affinity for said protein.

14           10. The method of Claim 9 wherein said antibody is  
15 conjugated with a toxic agent.

          11. Cell lines exhibiting defined amounts of DNA, RNA or  
protein specific for the v-erbB-related gene of claim 1.

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ABSTRACT OF DISCLOSURE

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The isolation, cloning and characterization of a human gene related to but distinct from EGF receptor gene has been described. Nucleotide sequence of the gene and amino acid sequence of the polypeptide encoded by the gene have been determined. The use of the nucleic acid probes and antibodies having specific binding affinity with said polypeptide for diagnostic and therapeutic purposes have also been described.